Utilising experimental NAMs to assess genomic instability in liver models





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Background

- Assessing genomic instability using traditional models may not necessarily be as predictive as we need them to be to replace and reduce in vivo testing of genotoxic and carcinogenic materials.
- The advantage of the human hepatocellular carcinoma (HepG2) cells are that when cultured as spheroids in a 3D format they can
 demonstrate liver-like functionality and metabolic activity.
- These crucial features can be used to support genotoxicity testing of chemicals, nanomaterials and pro-carcinogens.

overnight.

- Moreover, by including the use of the ultra-low attachment plates, these HepG2 spheroids can be grown, exposed, and then analysed in rapid time.
- The aim of this study was to establish a protocol to undertake genotoxicity testing via the in vitro alkaline comet assay using 3D HepG2 liver spheroids.

Methods



- Cells were seeded into ultra-low attachment (ULA) plates at 2500 cells/well and cultured for one week.
- The spheroids were treated for 24-hours with 300 μ M methyl-, and ethyl-methanesulfonate (MMS and EMS).
- Cytotoxicity was assessed before the spheroids were prepared for the *in vitro* alkaline comet assay. The average %Tail DNA intensity median was calculated using the Comet IV software. A total of 150 cells were scored per biological replicate (n=3)

Friday: Monday: Tuesday: Perform an acute (24-hour) exposure of Seed HepG2 cells and leave Harvest supernatant for endpoint analysis. Harvest spheroids for comet assay. for 48 hours to establish test material. spheroids. Tuesday: Dis-aggregate spheroids and Wednesday: Wednesday embed in low-melting point Unwind DNA, run electrophoresis tank at 1V/ Cells are stained with SYBR™ Gold. cm for 20 minutes. Remove slides and neutralise with PBS and 150 cells are then analysed for % Tail DNA Prepare 1 gel per exposure Intensity per biological replicate per condition. condition. Immerse slides in lysis buffer

Figure 1. HepG2 spheroid cell viability (A) and % Tail DNA Intensity (B) in the comet assay. The average % Tail DNA Intensity median was calculated for 150 cells per biological replicate, n=3.

Results



- Following a 24-hour exposure to MMS

 a statistically significant (p≤0.05)
 cytotoxic response was observed (A)
 with a 30% reduction in viable cells
 compared to the untreated spheroids.
- Statistically significant (p≤0.05) strand breaks (SBs) were observed using the comet assay in spheroids treated with both MMS and EMS (B) inducing 40% and 30% SBs respectively; indicative of a potent genotoxic response and in the case of MMS, correlating to a cytotoxic effect.

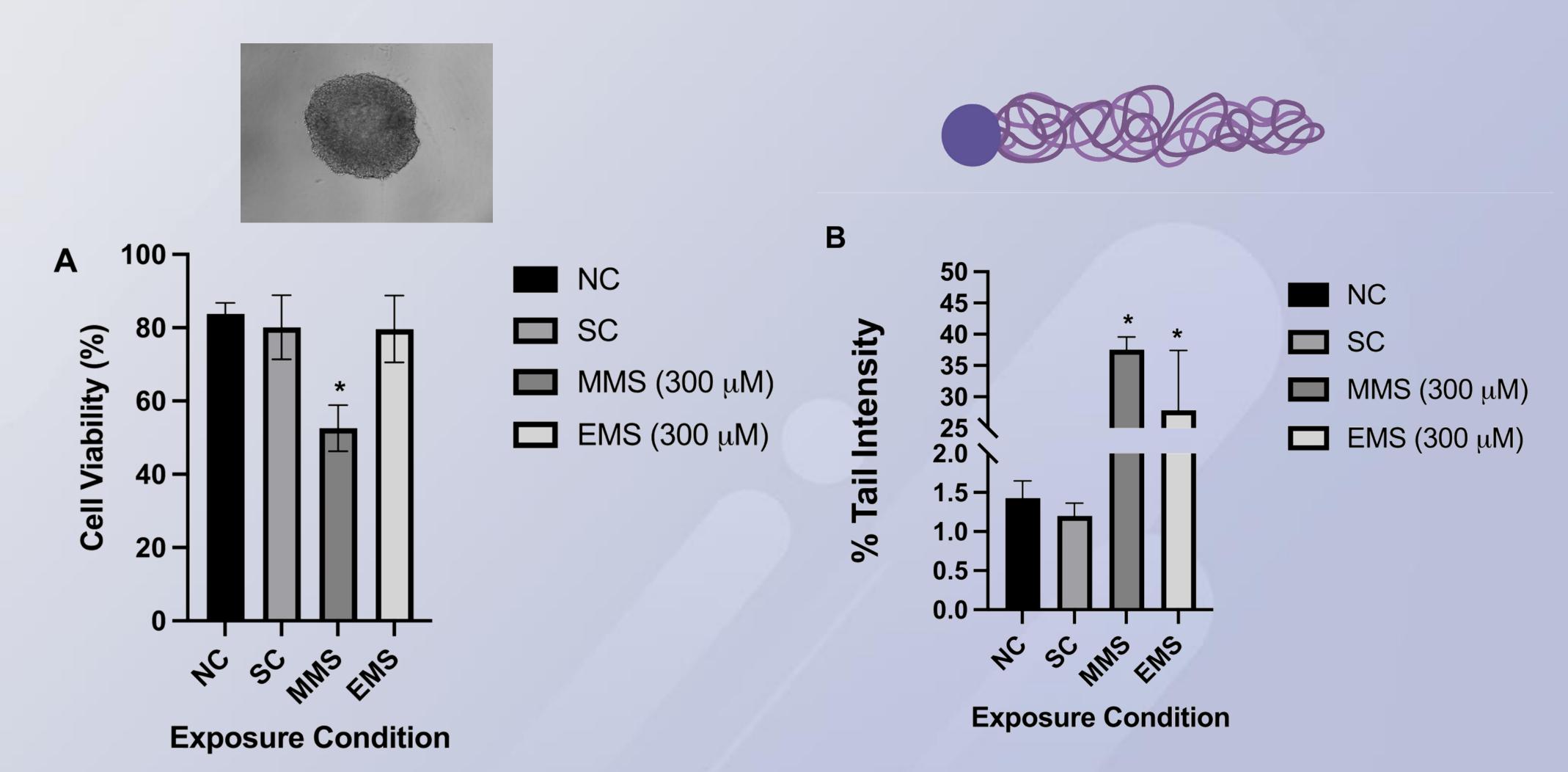


Figure 2. HepG2 spheroid cell viability (A) and % Tail DNA Intensity (B) in the comet assay. The average % Tail DNA Intensity median was calculated for 150 cells per biological replicate, n=3.





- Using the ULA plates improved the efficiency of generating HepG2 spheroids for genotoxicity evaluations.
- The in vitro alkaline comet assay could be readily applied for genotoxicity testing in this more complex in vitro test model.



